

Effect of Arbuscular Mycorrhizal Fungi on the Growth and Yield of Soybean (*Glycine max* L. Merrill) in Bauchi, Nigeria

Abstract

Soybeans (*Glycine max* L.) globally has been regarded as an economically important commodity that is highly traded and also a vital legume used as food source for both humans and animals. The objective of the study therefore is to determine the effect of arbuscular mycorrhizal fungi (AMF) on the growth and yield of soybean (*Glycine max* L.). The experiment was conducted in a screen house where two varieties of soybeans (TGX 1448 and TGX 1951) were grown in a 1 litre pods, filled with sterilized soil and three seeds were sown into each pod at a depth of 2 cm until germination, then reduced to one seedling. Different AMF dose (10 g, 20 g, 30 g, and 40 g) was inoculated at the time of seed sowing and non-inoculated pods as control (0 g). Various parameters were taken into consideration like plant height and number of leaves while shoot dry biomass, root dry biomass and yield attributes were taken at harvest. It was observed that the inoculated plants performed higher than the non-inoculated plants. Growth parameters such as plant height, number of leaves, shoot dry biomass, root dry biomass, and yield attributes increased with increase AMF dose. Therefore, it is concluded that AMF inoculation increase growth and yield of soybeans and can serve as biofertilizer.

Keywords: *Glycine max* L., arbuscular mycorrhizal fungi, inoculated, biofertilizer

Introduction

Soybeans (*Glycine max* L.) over the years have been regarded as one of the most economically important legumes in the food chain, with more than one fourth of the global population depending on it for food and other essentials such as animal feeds (Igiehon et al., 2021). The importance of soybean (*Glycine max* L.) as both oil crop and legumes to the food chain is paramount, belonging to the family Fabaceae (Adeyemi et al., 2020; Cera et al., 2017). Soybeans has 20% oil, when dry with other vital amount of minerals and vitamins present, and also, provides high quality protein for many households and processing industries in Nigeria (Omoigui et al., 2020).

Legumes have been an important way of earning for many farmers in most underdeveloped and developing nation, soybeans being known to improving soil fertility due to the ability to fix atmospheric nitrogen into the soil in the form that can be utilized by plants, thereby, lessening the need for organic and mineral fertilizers. In contrast, more than half of total nitrogen been added to the soil emanate from legumes – rhizobia symbiosis relationship (Bashir et al., 2022). Low yield in soybean farming are usually associated with nutrients imbalance, nutrients leaching and also, due to limited nutrients in the non-fertile soil (Bashir et al., 2022; Thuita et al., 2012)

Arbuscular Mycorrhiza fungi (AMF) form symbiosis relationship with most plant species by colonization of the host plant roots in order to source carbohydrates for their growth, development and continuous survival while in return, it provides minerals, nutrients and water to the host plant (Baslam & Goicoechea, 2012; Igiehon et al., 2021; Quiroga et al., 2019). A study by Sugiura et al., (2020) reveals that myristate can be a source of carbon and energy for AMF. Immediately the spore germinate, colonization of the plant root start by forming a hyphopodium on the surface of the root, which penetrate the rhizodermis through a pre-penetration apparatus, these colonizes root tissue intercellularly to form highly form arbuscules in cortical cells which the fungi uses to release minerals to the host plant (Etemadi et al., 2014; Harrison, 2012). Glomalin secreted by AMF help improves soil organic matter (SOM), soil structure, microbial activity, mitigate drought effects (Habibzabeh, 2015; Hong et al., 2018), bioremediation, and reduce loss of fertility (Mrabet et al., 2014; Priscila et al., 2021; Yang et al., 2017).

Several scholars have reported positive effects of using AMF to boost growth and yield of plants particularly legumes and cereals (Adeyemi et al., 2020; Buysens et al., 2016; Douds et al., 2016; Emam, 2016; Ming-hung et al., 2007; Novais et al., 2020; Zhu et al., 2010, 2014) and similarly, other scholars have reported no or less effects of using AMF (Farmer et al., 2007; Pellegrino & Bedini, 2014). Therefore, it is necessary to investigate the effect of different dose of AMF on the growth and yield of soybeans (*Glycine max* L.) varieties (TGX 1448 and TGX 1951).

Materials and Method

Experimental site

The experiment was conducted in the Screen house of Abubakar Tafawa Balewa University in Bauchi, Bauchi State, Nigeria.

Soil analysis

pH (H₂O), Electrical Conductivity (dsm⁻¹), Exchangeable Acidity (cmol kg⁻¹), Ca²⁺ (cmol kg⁻¹), Mg²⁺ (cmol kg⁻¹), K⁺ (cmol kg⁻¹), Na⁺ (cmol kg⁻¹), Cation Exchange Capacity (cmol kg⁻¹), Total Exchangeable Base (cmol kg⁻¹), Base Saturation (%), N (%), Organic Carbon (%), Organic Matter (%), Carbon to Nitrogen, Available Phosphorus (mg kg⁻¹), Clay, Sand, Silt, Texture were determined.

Source of Soybean seeds

Two varieties of soybean (TGX 1448 and TGX 1951) were purchased from Bauchi State Agricultural Development Programme (BASADP), Bauchi, Nigeria. Viability test was carried out according to

Source of AMF Inoculum

AMF inoculum was sourced from University of Aberdeen, School of Biological Sciences, Aberdeen, Scotland.

Screen House Experiment

Plants were grown in 1 litre pot with 4 replications. Different concentration of AMF *Glomus intraradices* (10g, 20g, 30g, and 40g) were inoculated and non-inoculated pot as control. Growth characteristics such as (plant height, number of leaves, root and shoot biomass, yield) were determined.

The pre-planting soil was collected in a transparent Ziplock polythene bag and taken to University of Maiduguri, Department of Soil Science for physical and chemical analysis. The result of the pre-planting soil analysis is presented in Table 1.

Experimental design and treatment

The experiment was set-up in a completely randomized design (CRD) using 1 litre pots filled with 5 mm sieved soil. Three soybean seeds were planted into each pot at a depth of 2 cm which after germination others were removed, maintaining one at each pot. The inoculation of AMF (10g, 20g, 30g, and 40g i.e. average of 40 spores per 10 g) were done at the time of sowing.

Data collection

Plant height, and number of leaves were recorded at two weeks intervals while Root biomass, shoot biomass, weight of seeds and number of seeds per plant were recorded at harvest i.e. at thirteen weeks.

Statistical analysis

Data collected from growth indices (plant height, number of leaves, root biomass, shoot biomass and yield) were analysed using DSAAT statistical software on Microsoft excel and GraphPad Prism version 8.0 software.

Result

The results of the physical and chemical properties of the experimental soil are presented in table 1. The result indicated that soil pH was 6.68 with EC of 0.84 (dsm^{-1}). Available phosphorus value of 12.25 (mg kg^{-1}) and the texture was identified as sandy loam.

Table 1. Physical and chemical properties of the pre-planting soil

Soil property	Value
pH (H_2O)	6.68
EC (dsm^{-1})	0.84
EA (cmol kg^{-1})	1.90
Ca ²⁺ (cmol kg^{-1})	2.40
Mg ²⁺ (cmol kg^{-1})	3.60
K ⁺ (cmol kg^{-1})	0.17
Na ⁺ (cmol kg^{-1})	0.13
CEC (cmol kg^{-1})	6.30
TEB (cmol kg^{-1})	8.20
Base Sat. (%)	76.83
N (%)	0.18
OC (%)	0.64

OM (%)	1.10
C:N	3.56
AP (mg kg ⁻¹)	12.25
Clay	16.50
Sand	75.30
Silt	8.20
Texture	Sandy loam

EC-Electrical conductivity, EA-Exchangeable acidity, Ca²⁺ -Calcium, Mg²⁺ -Magnesium, K⁺ - Potassium, Na⁺ -Sodium, CEC- Cation exchange capacity, TEB- Total exchangeable base, Base sat- Base saturation, N- Nitrogen, OC- Organic carbon, OM- Organic matter, C:N- Carbon to Nitrogen, and AP- Available Phosphorus.

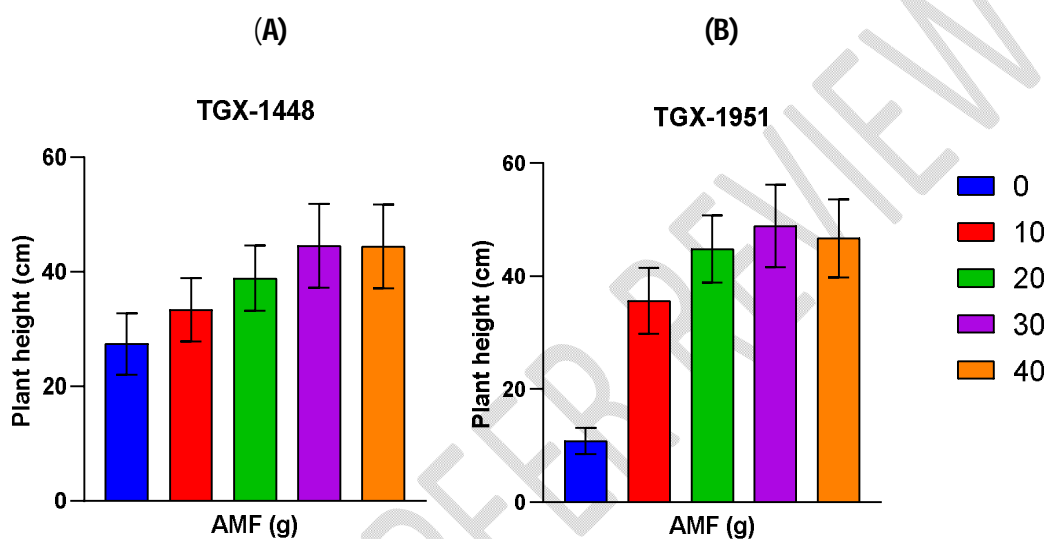


Figure 1: **A** - Effect of AMF (*Glomus intraradices*) on plant height of *Glycine max* (TGX 1448) **B** - Effect of AMF (*Glomus intraradices*) on plant height of *Glycine max* (TGX 1951)

Figure 1A shows the plant height of *Glycine max* variety TGX 1448 inoculated with different dose of arbuscular mycorrhiza fungi (AMF) of which at harvest, there is no significant difference between 30g and 40g AMF while 0 g i.e. control recorded less plant height and there is significant difference between control (0g), 10g and 20g AMF. Figure 1B shows plant height of *Glycine max* variety TGX 1951 inoculated with different dose of arbuscular mycorrhiza fungi (AMF), at harvest, 30 g AMF has the highest plant height but there is no significant difference between 20 g, 30 g and 40 g while there is significant difference between control (0g), 10g and 20g AMF.

(A)

(B)

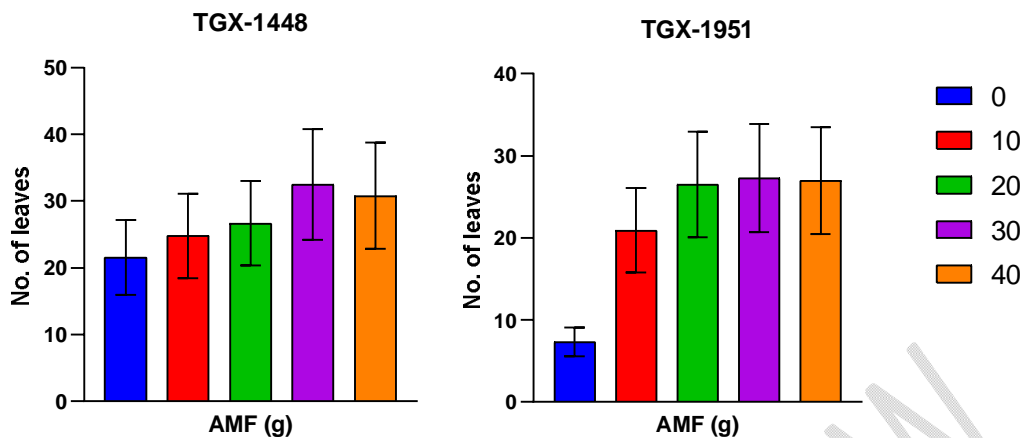


Figure 2: **A** -Effect of AMF (*Glomus intraradices*) on number of leaves of *Glycine max* (TGX 1448) **B** -Effect of AMF (*Glomus intraradices*) on number of leaves of *Glycine max* (TGX 1951)

Figure 2A shows number of leaves of *Glycine max* variety TGX 1448 inoculated with different dose of arbuscular mycorrhiza fungi (AMF), at harvest, 30g has the highest leaves number, but there is no significant difference between 30 g and 40 g AMF while there is no significant difference between control (0g), 10g and 20g respectively. Figure 2B shows number of leaves of *Glycine max* variety TGX 1951 inoculated with different dose of arbuscular mycorrhiza fungi (AMF) at harvest, there is no significant difference between 20 g, 30 g and 40 g while there is significant difference between control (0g), and 10g.

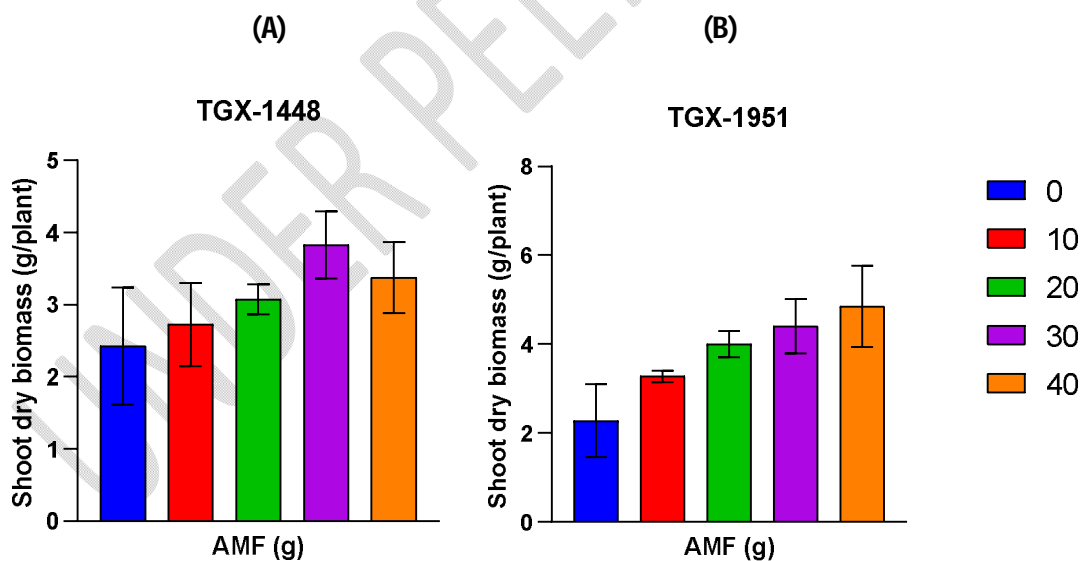


Figure 3: **A** - Effect of AMF (*Glomus intraradices*) on shoot dry biomass of *Glycine max* (TGX 1448) **B** - Effect of AMF (*Glomus intraradices*) on shoot dry biomass of *Glycine max* (TGX 1951)

Figure 3A shows shoot biomass of *Glycine max* (TGX 1448) inoculated with different dose of mycorrhizal (AMF). 30g has the highest weight, followed by 40g. For shoot, there is no significant difference between control (0 g), 10 g, 20 g and 40 g AMF. Figure 3B shows shoot

biomass of *Glycine max* (TGX 1951) inoculated with different dose of mycorrhizal (AMF). The weight increase with increased in AMF dose of which 40g has the highest weight in shoot biomass.

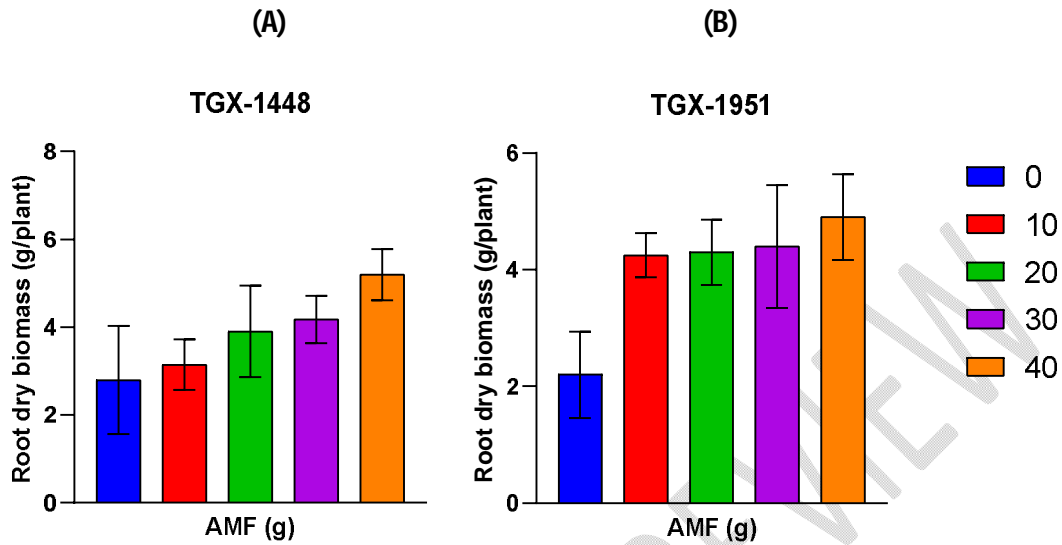


Figure 4 A - Effect of AMF (*Glomus intraradices*) on root dry biomass of *Glycine max* (TGX 1448) **B** - Effect of AMF (*Glomus intraradices*) on root dry biomass of *Glycine max* (TGX 1951)

Figure 4A shows root biomass of *Glycine max* (TGX 1448) inoculated with different dose of mycorrhizal (AMF). the weight increase with increased in AMF dose of which 40g has the highest root weight, while there is no significant difference between control, and 10g AMF. Subsequently, there is no significant difference between 20g, and 30g AMF. Figure 4B shows root biomass of *Glycine max* (TGX 1951) inoculated with different dose of mycorrhizal (AMF). In root biomass, the weight increase with increased in AMF dose of which 40g has the highest weight in root biomass while there is no significant difference between control, and 10g, 20 g and 30 g AMF. Subsequently, there is significant difference between non- mycorrhizal inoculated control (0g) with the inoculated mycorrhizal ones.

(A)

(B)

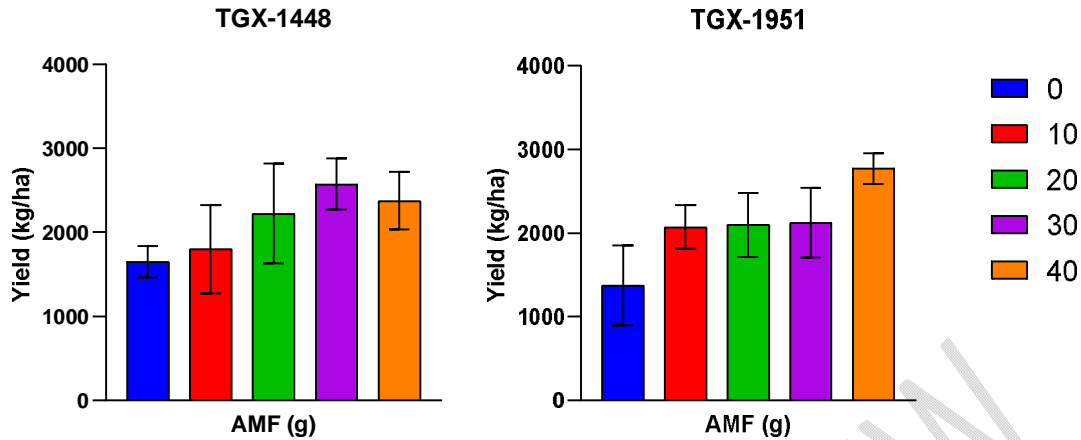


Figure 5: **A**- Effect of AMF (*Glomus intraradices*) on root dry biomass of *Glycine max* (TGX 1448) **B** - Effect of AMF (*Glomus intraradices*) on root dry biomass of *Glycine max* (TGX 1951)

Figure 5A shows yield of different varieties of *Glycine max* (TGX 1448) inoculated with different dose of AMF. Among the different treatments of variety TGX 1448, 30g has best yield as compared to the other treatments and there is no significant difference between control, and 20g, 30 g and 40 g AMF. Figure 5B shows yield of variety of *Glycine max* (TGX 1951) inoculated with different dose of AMF. Variety 2-TGX 1951 40g has the best yield as compared to the different treatments and there is no significant difference between 10 g, 20g, and 30 g AMF. Subsequently, there is significant difference between non- mycorrhizal inoculated control (0g) with the inoculated mycorrhizal ones.

Discussion

The used of AMF (*Glomus intraradices*) have increase plant growth and yield attributes of the two varieties (TGX 1448 and TGX 1951) of soybeans. These increase in overall growth and yield have earlier been reported by (Adeyemi et al., 2020; Klironomos et al., 2000). The presence of AMF has influence growth and yield attributes such as plant height and number of leaves due to the fact AMF may have influence nutrient uptake. AMF increase colonization in the rhizobia which subsequently provides the support by enhancing photosynthetic rate through the supply of phosphorus and nitrogen. The photosynthetic enzymes responsible for light harvesting complex solely depends on essentials nutrients supply by AMF of which phosphorus are known to stimulate many functions in plants such as canopy photosynthesis, nutrient movement and energy transfer in plants (Groot et al., 2003; Mo et al., 2016; Tereucán et al., 2022; Zhao et al., 2017). Similarly, AMF have positive effect on antioxidant enzyme activities in plants (Zhang et al., 2019)

The symbiotic relationship between plant and mycorrhiza positively enhance the root length, root biomass, root density, increase nutrient uptake especially nitrogen, phosphorus, iron and zinc (Barea, 2015; Delavaux et al., 2021; Ingraffia et al., 2019; Roupheal et al., 2015) and also, uptake of potassium (Ouledali et al., 2018; Wang et al., 2017). Root colonisation of the both varieties of soybeans (TGX 1448 and TGX 1951) inoculated with AMF were higher which may have resulted due to optimal value of available phosphorus in the soils (Carrasco et al., 2006;

Vasar et al., 2021). AMF symbiosis is more established predominantly in marginal soil allowing the secretion of root exudates by the plant which increased colonisation in the rhizobia(Adeyemi et al., 2020; Smith, 2008; Torrecillas et al., 2012).

Bacterial growth and vitality are influence by mycelial exudates which enhance the community of bacterial in the rhizosphere (Lindahl et al., 2007; Tanaka et al., 2022; Toljander et al., 2006). The ability of AMF hyphae to form symbiotic association with the plant root depends on the different bacterial groups present in the soil (Kohler et al., 2017; Scheublin et al., 2010). According to Tsoata et al., (2015) and Abdalla & Ahmed, (2021) AMF can be used as effective tool for ameliorating the negative impact of drought stress on plant by enhancing plant resistance and tolerance to abiotic stress which result in increased yield.

The plant height and number of leavesbetween the inoculated AMF plants and the non-inoculated AMF plants (0 g) in both soybeans varieties is visible, the inoculated plants performed higher than the non-inoculated AMF plants which agrees to the study of (Oliveira et al., 2022). Furthermore, shoot dry biomass and root dry biomass in the inoculated AMF plants exhibit similar pattern of performance with plant height and number of leaves of which the inoculated AMF plants have higher weight than the non-inoculated AMF plants which agrees with the study of (Oliveira et al., 2022).

In contrast, high yield performancerecorded in TGX 1951 at high AMF dose plants relates to increase growth parameters specifically plant height and number of leaves, of which high photosynthesis directly relates to increased assimilatory surface, resulting to increased shoot and root biomass and finally, increased yield attributes(Abdel et al., 2016; Adeyemi et al., 2020).

Conclusion

In conclusion, AMF (*Glomus intraradices*) exhibit greater potential to increased yield under favourable condition. The result validates the influence of AMF on the growth and yield of soybeans varieties (TGX 1448 and TGX 1951). It reveals that high yield attributes were observed with increase AMF dose in both varieties of soybeans which relates to high AMF colonisation around the roots, thereby increasing both water and nutrient uptake through the roots by the plant. TGX 1951 has the best performance with AMF inoculation then TGX 1448.

Abbreviations

AMF	Arbuscular mycorrhizal fungi
BASADP	Bauchi state agricultural development programme
CRD	Completely randomized design
DSAAT	Digital situational awareness assessment and training
SOM	Soil organic matter

References

- Abdalla, M., & Ahmed, M. (2021). Arbuscular Mycorrhiza Symbiosis Enhances Water Status and Soil- Plant Hydraulic Conductance Under Drought. *Frontiers in Plant Science*, 12(722954), 1–8. <https://doi.org/10.3389/fpls.2021.722954>
- Abdel, A., Abdel, H., Hashem, A., Rasool, S., Fathi, E., & Allah, A. (2016). Arbuscular Mycorrhizal Symbiosis and Abiotic Stress in Plants : A Review Arbuscular Mycorrhizal Symbiosis and Abiotic Stress in Plants : A Review. *J. Plant Biol.*, 59, 407–426. <https://doi.org/10.1007/s12374-016-0237-7>
- Adeyemi, N. O., Atayese, M. O., Olubode, A. A., & Akan, M. E. (2020). Effect of commercial arbuscular mycorrhizal fungi inoculant on growth and yield of soybean under controlled and natural field conditions. *Journal of Plant Nutrition*, 43(4), 487–499. <https://doi.org/10.1080/01904167.2019.1685101>
- Barea, J. M. (2015). Nutrient cycling in the mycorrhizosphere. *Journal of Soil Science and Plant Nutrition*, 25(2), 372–396.
- Bashir, M. (2022). EFFECTS OF SOIL TEXTURE AND NUTRIENTS APPLICATION ON SOYBEAN NUTRIENT UPTAKE, GROWTH AND YIELD RESPONSE. *Journal of Agriculture and Food Sciences*, 20(1), 227–241.
- Baslam, M., & Goicoechea, N. (2012). Water deficit improved the capacity of arbuscular mycorrhizal fungi (AMF) for inducing the accumulation of antioxidant compounds in lettuce leaves. *Mycorrhiza*, 22, 347–359. <https://doi.org/10.1007/s00572-011-0408-9>
- Buysens, C., César, V., Ferrais, F., Dupré, H., Boulois, D., & Declerck, S. (2016). Inoculation of Medicago sativa cover crop with Rhizophagus irregularis and Trichoderma harzianum increases the yield of subsequently-grown potato under low nutrient conditions. *Applied Soil Ecology*, 105, 137–143. <https://doi.org/10.1016/j.apsoil.2016.04.011>
- Carrasco, L., Caravaca, F., & Rolda, A. (2006). Stability of desiccated rhizosphere soil aggregates of mycorrhizal Juniperus oxycedrus grown in a desertified soil amended with a composted organic residue. *Soil Biology & Biochemistry*, 38, 2722–2730. <https://doi.org/10.1016/j.soilbio.2006.04.024>
- Cera, J. C., Streck, N. A., Augusto, C., & Fensterseifer, J. (2017). Soybean yield in future climate scenarios for the state of Rio Grande do Sul , Brazil. *Pesq. Agropec*, 52(6), 380–392. <https://doi.org/10.1590/S0100-204X2017000600002>
- Delavaux, C. S., Weigelt, P., Dawson, W., Essl, F., Kleunen, M. Van, König, C., Pergl, J., Py, P., Stein, A., Winter, M., Taylor, A., Schultz, P. A., Whittaker, R. J., Kreft, H., & Bever, J. D. (2021). Mycorrhizal types influence island biogeography of plants. *COMMUNICATIONS BIOLOGY*, 4, 1128. <https://doi.org/10.1038/s42003-021-02649-2>
- Douds, D. D., Wilson, D. O., Seidel, R., & Ziegler-ulsh, C. (2016). Scientia Horticulturae A method to minimize the time needed for formation of mycorrhizas in sweet corn seedlings for outplanting using AM fungus inoculum produced on-farm & . *Scientia Horticulturae*, 203, 62–68. <https://doi.org/10.1016/j.scienta.2016.03.015>
- Emam, T. (2016). Local soil , but not commercial AMF inoculum , increases native and non-native grass growth at a mine restoration site. *The Journal of the Society for Ecological Restoration*, 24(1), 35–44. <https://doi.org/10.1111/rec.12287>

- Etemadi, M., Gutjahr, C., Couzigou, J. M., Zouine, M., Laressergues, D., Timmers, A., Audran, C., Bouzayen, M., Bécard, G., & Combiér, J. P. (2014). Auxin perception is required for arbuscule development in arbuscular mycorrhizal symbiosis. *Plant Physiology*, *166*(1), 281–292. <https://doi.org/10.1104/pp.114.246595>
- Farmer, M. J., Li, X., Feng, G., Zhao, B., Chatagnier, O., & Gianinazzi, S. (2007). Molecular monitoring of field-inoculated AMF to evaluate persistence in sweet potato crops in China. *Applied Soil Ecology*, *35*, 599–609. <https://doi.org/10.1016/j.apsoil.2006.09.012>
- Groot, C. C. De, Boogaard, R. Van Den, Marcelis, L. F. M., & Harbinson, J. (2003). Contrasting effects of N and P deprivation on the regulation of photosynthesis in tomato plants in relation to feedback limitation. *Journal of Experimental Botany*, *54*(389), 1957–1967. <https://doi.org/10.1093/jxb/erg193>
- Habibzabeh, Y. (2015). Scientific members of Agricultural Research center of West Azarbaijan province, Urmia - Iran. *International Journal of Science*, *4*(3), 1–7.
- Harrison, M. J. (2012). Cellular programs for arbuscular mycorrhizal symbiosis. *Current Opinion in Plant Biology*, *15*(6), 691–698. <https://doi.org/10.1016/j.pbi.2012.08.010>
- Hong, N., Csintalan, Z., & Posta, K. (2018). Plant Physiology and Biochemistry Arbuscular mycorrhizal fungi mitigate negative effects of combined drought and heat stress on tomato plants. *Plant Physiology and Biochemistry*, *132*(September), 297–307. <https://doi.org/10.1016/j.plaphy.2018.09.011>
- Igiehon, N. O., Babalola, O. O., Cheseto, X., & Torto, B. (2021). Effects of rhizobia and arbuscular mycorrhizal fungi on yield, size distribution and fatty acid of soybean seeds grown under drought stress. *Microbiological Research*, *242*, 126640. <https://doi.org/10.1016/j.micres.2020.126640>
- Ingraffia, R., Id, G. A., Frenda, A. S., & Id, D. G. (2019). Impacts of arbuscular mycorrhizal fungi on nutrient uptake, N₂ fixation, N transfer, and growth in a wheat / faba bean intercropping system. *PLoS ONE*, *14*(3), 1–16.
- Klironomos, J. N., McCune, J., Miranda, H. A., & Neville, J. (2000). The influence of arbuscular mycorrhizae on the relationship between plant diversity and productivity. *Ecology Letters*, *3*, 137–141. <https://doi.org/10.1046/j.1461-0248.2000.00131.x>
- Kohler, J., Roldán, A., Campoy, M., & Caravaca, F. (2017). Unraveling the role of hyphal networks from arbuscular mycorrhizal fungi in aggregate stabilization of semiarid soils with different textures and carbonate contents. *Plant and Soil*, *410*, 273–281. <https://doi.org/10.1007/s11104-016-3001-3>
- Lindahl, D., Paul, L. R., Elfstrand, M., Finlay, R. D., & Toljander, J. F. (2007). Influence of arbuscular mycorrhizal mycelial exudates on soil bacterial growth and community structure. *FEMS Microbiol Ecol*, *61*, 295–304. <https://doi.org/10.1111/j.1574-6941.2007.00337.x>
- Ming-hung, A. L. X. Z., Lou, W. Z. Y. L., & Zhu, Y. W. Y. (2007). Increase of multi-metal tolerance of three leguminous plants by arbuscular mycorrhizal fungi colonization. *Environ Geochem Health*, *29*, 473–481. <https://doi.org/10.1007/s10653-007-9116-y>
- Mo, Y., Wang, Y., Yang, R., Zheng, J., Liu, C., & Li, H. (2016). Regulation of Plant Growth,

Photosynthesis , Antioxidation and Osmosis by an Arbuscular Mycorrhizal Fungus in Watermelon Seedlings under Well-Watered and Drought Conditions. *Frontiers in Plant Science*, 7(644), 1–15. <https://doi.org/10.3389/fpls.2016.00644>

- Mrabet, S. El, Ouahmane, L., & Mousadik, A. El. (2014). The Effectiveness of Arbuscular Mycorrhizal Inoculation and Bio-Compost The Effectiveness of Arbuscular Mycorrhizal Inoculation and Bio-Compost Addition for Enhancing Reforestation with *Argania spinosa* in Morocco. *Open Journal of Forestry*, 4(1), 14–23. <https://doi.org/10.4236/ojf.2014.41003>
- Novais, C. B. De, Sbrana, C., Jesus, C., Felicianus, L., Rouws, M., Giovannetti, M., Avio, L., Siqueira, J. O., José, O., & Júnior, S. (2020). Mycorrhizal networks facilitate the colonization of legume roots by a symbiotic nitrogen-fixing bacterium. *Mycorrhiza*, 30, 389–396.
- Oliveira, T. C., Silva, J., Cabral, R., Santana, L. R., Tavares, G. G., Dionísio, L., Santos, S., Paim, T. P., Müller, C., Silva, F. G., Costa, A. C., Souchie, E. L., & Mendes, G. C. (2022). The arbuscular mycorrhizal fungus *Rhizophagus clarus* improves physiological tolerance to drought stress in soybean plants. *Scientific Reports*, 12(9044), 1–15. <https://doi.org/10.1038/s41598-022-13059-7>
- Omoigui, L. O., Kamara, A. Y., Kamai, N., Dugje, I. Y., Ekeleme, F., Kumar, P. L., Ademulegun, T., & Solomon, R. (2020). *Guide to Soybean Production in Northern Nigeria Guide to Soybean Production in Northern*. International Institute of Tropical Agriculture (IITA) Ibadan, Nigeria.
- Ouledali, S., Ennajeh, M., Zrig, A., Gianinazzi, S., & Khemira, H. (2018). Estimating the contribution of arbuscular mycorrhizal fungi to drought tolerance of potted olive trees (*Olea europaea*). *Acta Physiologiae Plantarum*, 40(5), 1–13. <https://doi.org/10.1007/s11738-018-2656-1>
- Pellegrino, E., & Bedini, S. (2014). Soil Biology & Biochemistry Enhancing ecosystem services in sustainable agriculture : Biofertilization and biofortification of chickpea (*Cicer arietinum* L.) by arbuscular mycorrhizal fungi. *Soil Biology and Biochemistry*, 68, 429–439. <https://doi.org/10.1016/j.soilbio.2013.09.030>
- Priscila, M., Cristiane, da S., Júnior, D., Carlos, C., Marcos, P., & Everaldo, Z. (2021). Beneficial services of Glomalin and Arbuscular Mycorrhizal fungi in degraded soils in Beneficial services of Glomalin and Arbuscular Mycorrhizal fungi in degraded soils in. *Scientia Agricola*, 79(5), 1–14. <https://doi.org/10.1590/1678-992X-2021-0064>
- Quiroga, G., Erice, G., Aroca, R., Chaumont, F., & Ruiz-lozano, J. M. (2019). Contribution of the arbuscular mycorrhizal symbiosis to the regulation of radial root water transport in maize plants under water deficit. *Environmental and Experimental Botany*, 167(May), 103821. <https://doi.org/10.1016/j.envexpbot.2019.103821>
- Rouphael, Y., Franken, P., Schneider, C., Schwarz, D., Giovannetti, M., Agnolucci, M., Pascale, S. De, Bonini, P., & Colla, G. (2015). Scientia Horticulturae Arbuscular mycorrhizal fungi act as biostimulants in horticultural crops. *Scientia Horticulturae*, 196, 91–108. <https://doi.org/10.1016/j.scienta.2015.09.002>
- Scheublin, T. R., Sanders, I. R., Keel, C., & Meer, J. R. Van Der. (2010). Characterisation of

- microbial communities colonising the hyphal surfaces of arbuscular mycorrhizal fungi. *The ISME Journal*, 4, 752–763. <https://doi.org/10.1038/ismej.2010.5>
- Smith, J. E. (2008). Mycorrhizal Symbiosis (Third Edition). In *Academic Press, New York* (Issue 6). <https://doi.org/10.2136/sssaj2008.0015br>
- Sugiura, Y., Akiyama, R., Tanaka, S., Yano, K., Kameoka, H., & Marui, S. (2020). Myristate can be used as a carbon and energy source for the asymbiotic growth of arbuscular mycorrhizal fungi. *PNAS*, 117(41), 25779–25788. <https://doi.org/10.1073/pnas.2006948117>
- Tanaka, S., Hashimoto, K., Kobayashi, Y., Yano, K., Maeda, T., Kameoka, H., Ezawa, T., Saito, K., Akiyama, K., & Kawaguchi, M. (2022). Asymbiotic mass production of the arbuscular mycorrhizal fungus *Rhizophagus clarus*. *COMMUNICATIONS BIOLOGY*, 5(43), 1–9. <https://doi.org/10.1038/s42003-021-02967-5>
- Tereucán, G., Ruiz, A., Nahuelcura, J., Oyarzún, P., Santander, C., Winterhalter, P., Ademar, P., Ferreira, A., & Cornejo, P. (2022). Shifts in biochemical and physiological responses by the inoculation of arbuscular mycorrhizal fungi in *Triticum aestivum* growing under drought conditions. *Society of Chemical Industry*, 102, 1927–1938. <https://doi.org/10.1002/jsfa.11530>
- Thuita, M., Pypers, P., & Herrmann, L. (2012). Commercial rhizobial inoculants significantly enhance growth and nitrogen fixation of a promiscuous soybean variety in Kenyan soils. *Biol Fertil Soils*, 48(January), 87–96. <https://doi.org/10.1007/s00374-011-0611-z>
- Toljander, J. F., Artursson, V., Paul, L. R., Jansson, J. K., & Finlay, R. D. (2006). Attachment of different soil bacteria to arbuscular mycorrhizal fungal extraradical hyphae is determined by hyphal vitality and fungal species. *FEMS Microbiol Lett*, 254, 34–40. <https://doi.org/10.1111/j.1574-6968.2005.00003.x>
- Torrecillas, E., Alguacil, M. M., & Roldán, A. (2012). Host Preferences of Arbuscular Mycorrhizal Fungi Colonizing Annual Herbaceous Plant Species in Semiarid Mediterranean Prairies. *Applied and Environmental Microbiology*, 78(17), 6180–6186. <https://doi.org/10.1128/AEM.01287-12>
- Tsoata, E., Njock, S. R., Youmbi, E., & Nwaga, D. (2015). Early effects of water stress on some biochemical and mineral parameters of mycorrhizal *Vigna subterranea* (L.) Verdc. (Fabaceae) cultivated in Cameroon. *International Journal of Agronomy and Agricultural Research (IJAAAR)*, 7(2), 21–35.
- Vasar, M., Davison, J., Sepp, S., Öpik, M., Moora, M., Koorem, K., Meng, Y., Oja, J., Akhmetzhanova, A. A., Al-quraishy, S., Onipchenko, V. G., Cantero, J. J., Glassman, S. I., Hozzein, W. N., & Zobel, M. (2021). Arbuscular Mycorrhizal Fungal Communities in the Soils of Desert Habitats. *Microorganisms*, 9(229), 1–14.
- Wang, W., Shi, J., Xie, Q., Jiang, Y., Yu, N., & Wang, E. (2017). Nutrient Exchange and Regulation in Arbuscular Mycorrhizal Symbiosis. *Molecular Plant*, 10(9), 1147–1158. <https://doi.org/10.1016/j.molp.2017.07.012>
- Yang, Y., He, C., Huang, L., Ban, Y., & Tang, M. (2017). The effects of arbuscular mycorrhizal

fungi on glomalin-related soil protein distribution , aggregate stability and their relationships with soil properties at different soil depths in lead- zinc contaminated area. *PLOS ONE*, 12(8), 1–19.

Zhang, Z., Zhang, J., Xu, G., Zhou, L., & Li, Y. (2019). Arbuscular mycorrhizal fungi improve the growth and drought tolerance of *Zenia insignis* seedlings under drought stress. *New Forests*, 50(4), 593–604. <https://doi.org/10.1007/s11056-018-9681-1>

Zhao, X., Chen, T., Feng, B., Zhang, C., & Peng, S. (2017). Non-photochemical Quenching Plays a Key Role in Light Acclimation of Rice Plants Differing in Leaf Color. *Frontiers in Plant Science*, 7(January), 1–17. <https://doi.org/10.3389/fpls.2016.01968>

Zhu, X., Song, F., & Xu, H. (2010). Influence of arbuscular mycorrhiza on lipid peroxidation and antioxidant enzyme activity of maize plants under temperature stress. *Mycorrhiza*, 20, 325–332. <https://doi.org/10.1007/s00572-009-0285-7>

Zhu, X., Song, F., & Xu, H. (2014). Arbuscular mycorrhizae improves low temperature stress in maize via alterations in host water status and photosynthesis Arbuscular mycorrhizae improves low temperature stress in maize via alterations in host water status and photosynthesis. *Plant Soil*, 331(June 2010), 129–137. <https://doi.org/10.1007/s11104-009-0239-z>

UNDER PEER REVIEW